

difficult matrices is described.

# **Rapid Extraction, Cleanup and Determination of Multiple Pesticide Residues in Difficult Matrices Utilizing** Energized Dispersive Extraction and UPLC MS/MS

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### Introduction

In the modern world, consumers increasingly want to know what is in their food and that

the substances they are putting in their body are safe. This, along with stringent

regulatory requirements, is leading the call for improved extraction of food contaminates

such as pesticides. The QuEChERS method has been shown to be practical for pesticide analysis on a number of different sample types. Some matrices, such as avocado or hops, can be very difficult to work with and their extraction is a manual and tedious process that is a bottleneck. An alternative to the OuEChERS method that both helps improve pesticide recoveries in difficult matrices while also offering a more rapid and efficient process would be beneficial to the modern world. A new sample preparation technology, Energized Dispersive Extraction, paired with UPLC MS/MS analysis that allows for extraction, cleanup, filtration and analysis in less than fifteen minutes of

# **UPLC** Conditions

• Waters Acquity UPLC BEH C18 1.7 µm 2.1 x 50 mm column

#### 5 µL injections

- Mobile Phase A: 10 mM Ammonium Acetate in Water
- Mobile Phase B: 5 mM Ammonium Acetate in Methanol

| Time (min) | Flow Rate (mL/min) | %A | %В |
|------------|--------------------|----|----|
| Initial    | 0.25               | 95 | 5  |
| 2          | 0.25               | 95 | 5  |
| 6          | 0.25               | 60 | 40 |
| 12         | 0.25               | 10 | 90 |
| 14         | 0.25               | 10 | 90 |
| 16         | 0.25               | 95 | 5  |

· Quantitation was based on a 7 point multi-level calibration curve using Multiple Reactions Monitoring

## MS/MS Conditions

Canadian Pesticide Mix 4 in LCMS Acetonitrile

| Name                  | Transition | Cone Voltage (V) |
|-----------------------|------------|------------------|
| Acephate              | 143        | 16               |
| Chlorpyriphos         | 97         | 24               |
| Coumaphos             | 227        | 50               |
| Diazinon              | 169        | 24               |
| Dichlorvos            | 109        | 34               |
| Dimethoate            | 199        | 30               |
| Prophos               | 131        | 36               |
| Etofenprox            | 177        | 60               |
| Etoxazole             | 141        | 54               |
| Terrazole             | 105        | 52               |
| Fensulfothion         | 281        | 32               |
| Fenthion              | 169        | 32               |
| Malathion             | 127        | 30               |
| Methyl parathion      | 125        | 36               |
| Mevinphos             | 127        | 26               |
| Imidan (Phosmet)      | 160        | 30               |
| Spiroxamine           | 144        | 36               |
| Tetrachlorvinphos (Z) | 127        | 42               |
| Thiophanate-methyl    | 151        | 28               |

| Pesticide Recovery Data |                |             |  |  |  |
|-------------------------|----------------|-------------|--|--|--|
| Name                    | Recovery (n=3) | % RSD (n=3) |  |  |  |
| Acephate                | 136            | 16          |  |  |  |
| Chlorpyriphos           | 98             | 24          |  |  |  |
| Coumaphos               | 98             | 50          |  |  |  |
| Diazinon                | 103            | 24          |  |  |  |
| Dichlorvos              | 97             | 34          |  |  |  |
| Dimethoate              | 94             | 30          |  |  |  |
| Prophos                 | 121            | 36          |  |  |  |
| Etofenprox              | 103            | 60          |  |  |  |
| Etoxazole               | 114            | 54          |  |  |  |
| Terrazole               | 103            | 52          |  |  |  |
| Fensulfothion           | 64             | 32          |  |  |  |
| Fenthion                | 100            | 32          |  |  |  |
| Malathion               | 88             | 30          |  |  |  |
| Methyl parathion        | 96             | 36          |  |  |  |
| Mevinphos               | 107            | 26          |  |  |  |
| Imidan (Phosmet)        | 93             | 30          |  |  |  |
| Spiroxamine             | 88             | 36          |  |  |  |
| Tetrachlorvinphos (Z)   | 95             | 42          |  |  |  |
| Thiophanate-methyl      | 117            | 28          |  |  |  |

16 out of 19 compounds with %RSD values of 7 or less

The three compounds with higher %RSD values are most likely due to matrix interference.

1000

1200

- MS System: Waters Acquity H Class, Xevo TQD
- · Ionization Mode: ESI Capillary Voltage: 0.10kV
- Source temp: 150°C

100.000 50.000 0

Probe temp: 600°C

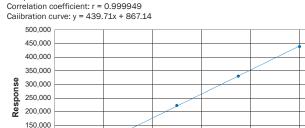
Sampling Rage: 10Hz

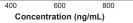
### **Pesticide Analysis**

Matrix Match Hops Calibration Curve

Compound name: Diazinon

200

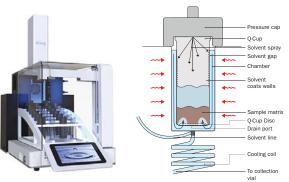




# Conclusions

The new energized dispersive extraction method for the extraction of pesticides from difficult food matrices vielded comparable or better recoveries compared to OuEChERS. Furthermore; the new energized dispersive method offered faster run times, and a automated simplified approach compared to alternative methods. Combined with a rapid UPLC MS/MS method, extraction and analysis of the samples is completed in under fifteen minutes. Energized dispersive extraction offers a good and economical option for the extraction of pesticides from all types of food matrices.

## **Energized Dispersive Extraction**



Q-Cup Technology combines the processes of Pressurized Fluid Extraction and Dispersive Solid Phase Extraction into one instrument.

### **Extraction of Pesticides**

### **EDGE Method**

### Sample extraction and sample clean up together

1. Add dSPE sorbent (same material can be used as in cleanup of the QuEChERS method) and transfer 15 g homogenized food sample to Q-Cup

- 2. Place Q-Cup in the EDGE
- 3. Run the 5 min EDGE method: 30 mL extraction at 100 °C
- 4. Transfer extract to a vial for concurrent analysis

### AOAC 2007.01 Method Procedure

### Sample extraction

- 1. Transfer 15 g homogenized sample to 50 mL centrifuge tube
- 2. Add 15 mL 1% acetic acid in acetonitrile + 1.5 g NaAc + 6 g MgSO<sub>4</sub>
- 3. Shake vigorously 1 min
- 4. Centrifuge > 1500 U/min for 1 min

### Sample Cleanup

1. Transfer 1-8 mL of acetonitrile layer to tube with 150 mg  $MgSO_4$  + 50 mg PSA per mL extract

- 2. Shake vigorously 30 sec
- 3. Centrifuge > 1500 U/min for 1 min
- 4. Transfer supernatant to a vial for concurrent analysis

# **Hops Extracts**

